



Distribution and genotypic analysis of *Enterocytozoon bieneusi* from cats in Korea



Heon-Moo Park¹ , Haeseung Lee² , So-Young Sung¹ , Kaifa Nazim³ , Bo-Yoon Jang⁴ , Ki-Chang Sung⁴ ,
Seung-Hun Lee⁵ , Min-Goo Seo^{1,6} , Man Hee Rhee^{1,6} , Dongmi Kwak^{1,6,*}

¹Department of Veterinary Medicine, College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea;

²Veterinary Epidemiology Division, Animal and Plant Quarantine Agency, Gimcheon 39660, Korea; ³Department of Veterinary Parasitology, Khalsa College of Veterinary & Animal Sciences, Punjab 143001, India; ⁴Department of Animal Health Management, Daegu Health College, Daegu 41453, Korea; ⁵Department of Veterinary Medicine, College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea; ⁶Institute for Veterinary Biomedical Science, College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea

Abstract

Received: 24 March 2025

Accepted: 7 May 2025

*Correspondence

dmiwak@knu.ac.kr

Citation

Park HM, Lee H, Sung SY, Nazim K,
Jang BY, Sung KC, Lee SH, Seo MG,
Rhee MH, Kwak D.

Distribution and genotypic analysis of
Enterocytozoon bieneusi from cats in
Korea.

Parasites Hosts Dis 2025;63(2):188–194.

Enterocytozoon bieneusi is an opportunistic microsporidian parasite with zoonotic potential that causes gastrointestinal illness in humans and animals. This study aimed to investigate the presence and genetic diversity of *E. bieneusi* from cats in Korea and to assess the potential public health risks associated with zoonotic genotypes. Among the 137 feline fecal samples, 4 (2.9%) were PCR-positive for *E. bieneusi*. In addition, 2 *E. bieneusi* genotypes were identified: Type IV, a known zoonotic genotype belonging to Group 1, and KCAT1, a novel genotype with zoonotic potential belonging to Group 1. This study is the first to report on these genotypes from cats in Korea, most of which were companion cats visiting veterinary clinics. Despite the low detection rate, the presence of zoonotic genotypes in companion cats is a potential public health concern because of the close physical interaction between cats and their human caregivers. These findings indicate the importance of routine monitoring and the molecular characterization of *E. bieneusi* in companion animals to comprehensively understand their zoonotic transmission patterns and to guide future risk assessments and preventive strategies.

Keywords: *Enterocytozoon bieneusi*, zoonosis, genotype, cat

The phylum Microsporidia includes obligate unicellular parasites that infect various vertebrate and invertebrate hosts, with *Enterocytozoon bieneusi* being the most common species found in humans [1]. Over 90% of human microsporidiosis cases are due to *E. bieneusi* [2], which causes opportunistic infections, especially in immunocompromised patients, such as those with acquired immune deficiency syndrome and cancer, as well as those who have undergone organ transplantation [3,4]. *E. bieneusi* cannot perform metabolic processes outside the host cell, and it can only survive externally in the spore stage. Upon ingestion by a suitable host, the spore invades the intestinal cells, causing infection. Mature spores are excreted through feces, and the life cycle of *E. bieneusi* continues through contact with infected animals or humans or via consumption of contaminated water or food in the environment [5].

Enterocytozoon bieneusi spores are oval and are among the smallest Microsporidia (0.70–0.98 $\mu\text{m} \times 1.08$ –1.64 μm) [5], making microscopic diagnosis difficult and requiring expertise. Therefore, PCR-based genotyping, which mainly targets the ribosomal internal transcribed spacer (ITS) region, is widely used to identify genetic characteristics and diver-

© 2025 The Korean Society for
Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Author contributions

Conceptualization: Kwak D
 Data curation: Lee H, Kwak D
 Formal analysis: Park HM
 Investigation: Park HM, Lee H
 Methodology: Park HM, Lee H, Sung SY
 Project administration: Kwak D
 Supervision: Kwak D
 Validation: Nazim K, Jang BY, Sung KC,
 Lee SH, Seo MG, Rhee MH
 Visualization: Park HM, Lee H, Nazim K
 Writing- original draft: Park HM, Lee H,
 Sung SY
 Writing- review and editing: Nazim K,
 Jang BY, Sung KC, Lee SH, Seo MG,
 Rhee MH, Kwak D

Conflict of interest

Dongmi Kwak serves as an editor of *Parasites, Hosts and Diseases* but had no involvement in the decision to publish this article. No other potential conflicts of interest relevant to this study were reported.

ORCID

Heon-Moo Park
<https://orcid.org/0009-0004-4751-4866>
 Haeseung Lee
<https://orcid.org/0000-0002-9587-5488>
 So-Young Sung
<https://orcid.org/0009-0002-0290-7696>
 Kaifa Nazim
<https://orcid.org/0000-0002-1524-3035>
 Bo-Yoon Jang
<https://orcid.org/0009-0003-2630-0044>
 Ki-Chang Sung
<https://orcid.org/0009-0001-2483-8882>
 Seung-Hun Lee
<https://orcid.org/0000-0002-6244-0381>
 Min-Goo Seo
<https://orcid.org/0000-0003-1752-5105>
 Man Hee Rhee
<https://orcid.org/0000-0002-3088-1318>
 Dongmi Kwak
<https://orcid.org/0000-0003-0876-3179>

sity [6]. To date, >685 *E. bieneusi* genotypes are widely distributed across more than 11 genotypic groups. Group 1 being the largest includes many genotypes found in humans and animals [1,5,7], such as genotypes D and Type IV, which are identified in several countries, including Korea [6].

In Korea, *E. bieneusi* has been studied in industrial animals (cattle, pigs, and horses) and in wild animals (bats, raccoon dogs, deer, leopard cats, wild boars, and wild deer) [8–10]. The *E. bieneusi* genotypes that are confirmed in Korea include CAF1, CEBd, D, H, hores1, KBAT3, Peru11, PigEBITS3 to PigEBITS5, and Type IV (group 1); BEB8, CEBa, CEBf, I, J, KBAT1, KBAT2, and KBAT4 (group 2); and horse2 (group 6) [6,9,11]. However, only few studies have investigated the prevalence of *E. bieneusi* in Korea, with only 1 study focusing on cats living in shelters on Jeju Island [11]. Jeju Island is located approximately 80 km from the southern coast of the Korean Peninsula. Given its geographical isolation and volcanic origins, the region exhibits a climate and natural environment that are markedly distinct from those of the mainland. The presence of zoonotic genotypes in companion cats could pose a public health concern because of the close interaction between companion cats and their human owners. Moreover, shelter cats may come into contact with the shelter staff during their temporary stay and with new owners during adoption. Therefore, this study aimed to assess the prevalence of *E. bieneusi* in cats, including companion and shelter cats, in Korea and to determine its public health implications.

From January 2017 to November 2022, veterinarians collected 137 feline fecal samples from animal clinics and shelters across Korea. Fresh fecal samples were placed in individual tubes, transported in ice-packed boxes at 4°C, and transported to the laboratory for DNA extraction. Sample information (including age, sex, season, region, the presence or absence of diarrhea, and animal source) was recorded, with unclear or questionable data labeled as “unknown.” Age was categorized into 5 groups: kittens (<6 months), juniors (<2 years), adults (<10 years), seniors (≥10 years), and unknown age. The sampling region was divided into northern (Gangwon and Gyeonggi), central (South Chungcheong, North Chungcheong, North Gyeongsang, and North Jeolla), and southern (South Gyeongsang, South Jeolla, and Jeju) regions in accordance with the administrative district boundaries.

Fecal samples from cats were collected in accordance with the project guidelines by veterinarians at local, government-run veterinary institutes during monitoring, surveillance, treatment, or regular medical checkups after the receipt of oral consent from the cat owners. Fecal collection did not cause any harm to the animals and did not require ethical approval.

DNA was extracted using a QIAamp Fast DNA Stool Mini Kit (Qiagen, Hilden, Germany) in accordance with the manufacturer’s instructions. DNA quantity and quality were evaluated using the Infinite 200 PRO NanoQuant plate reader (Tecan, Mannedorf, Switzerland). In addition, PCR amplification was performed using AccuPower HotStart PCR Premix (Bioneer, Daejeon, Korea), and nested PCR was performed to amplify the *E. bieneusi* ITS region for detection [12,13]. For the first round of PCR, ITSf1 (5′-GGTCATAGGGA-TGAAGAG-3′) and ITSr1 (5′-TTCGAGTTCTTTCGCGCTC-3′) were used as the forward and reverse primers, respectively. For the second round of PCR, ITSf2 (5′-GCTCT-GAATATCTATGGCT-3′) and ITSr2 (5′-ATCGCCGACGGATCCAAGTG-3′) were used as the forward and reverse primers, respectively [14]. PCR amplification was performed

using Mastercycler Pro (Eppendorf, Hamburg, Germany) under the following conditions: initial denaturation at 95°C for 5 min, followed by 35 cycles of denaturation at 95°C for 30 sec, annealing at 55°C for 30 sec, and extension at 72°C for 30 sec, with a final extension at 75°C for 5 min.

Among the 137 collected specimens, 4 were positive for *E. bieneusi* (Table 1), with a prevalence of 2.9%. This value was lower than that reported for other animals, such as cats in Korea (Jeju Island, 3.8%) [11], wild animals (45.2%), calves (16.9%), cattle (14.9%), and pigs (14.2%) [8], with only bats and wild deer showing lower prevalences of 2.6% and 6.8%, respectively [8,10]. Based on these studies, the prevalence rate of *E. bieneusi* varies according to animal type, sample size, and geographical region. This variation was also observed in previous research on cats [1,11,15].

In this study, the prevalence of *E. bieneusi* was analyzed according to region, season, sex, age, presence of diarrhea, and animal source. For statistical analysis, the chi-square test was performed using the IBM SPSS Statistics version 26 (IBM Corp., Armonk, NY, USA), and the results indicated no significant risks.

When comparing by region, the prevalence of *E. bieneusi* was 3.1% and 5.6% in the northern and central regions, respectively. With regard to season, the prevalence of *E. bieneusi* was 6.3% in summer, 3.6% in autumn, and 2.1% in winter. In addition, the prevalence of *E. bieneusi* was highest in females (3.4%), followed by males (2.8%) and those of unknown sex (2.3%). With regard to age, the prevalence of *E. bieneusi* was 4.5% in kittens aged 6 months, 4.2% in juniors aged <2 years old, and 2.3% in cats of unknown age. The *E. bieneusi* ITS region was not detected in diarrheal stools, but it occurred with a prevalence of 3.4% in normal stools. Finally, the prevalence of *E. bieneusi* was 3.1% in animal samples

Table 1. Prevalence of *Enterocytozoon bieneusi* in cats in Korea

Variable		No. of samples tested	No. of samples positive for <i>E. bieneusi</i> (%)	P-value
Region	Northern	65	2 (3.1)	0.626
	Central	36	2 (5.6)	
	Southern	31	0	
	Unknown	5	0	
Season	Spring	18	0	0.696
	Summer	16	1 (6.3)	
	Autumn	56	2 (3.6)	
	Winter	47	1 (2.1)	
Sex	Female	58	2 (3.4)	1.000
	Male	36	1 (2.8)	
	Unknown	43	1 (2.3)	
Age	Kitten (<6 months)	44	2 (4.5)	0.927
	Junior (<2 years)	24	1 (4.2)	
	Adult (<10 years)	22	0	
	Senior (>11 years)	3	0	
	Unknown	44	1 (2.3)	
Fecal type	Diarrhea	18	0	1.000
	Normal	119	4 (3.4)	
Animal source	Clinic	98	3 (3.1)	1.000
	Shelter	39	1 (2.6)	
Total		137	4 (2.9)	

collected from clinics and 2.6% in samples obtained from shelters (Table 1). Given the limited number of samples tested in each region and season, the association between the prevalence of *E. bieneusi* and specific geographical regions were not confirmed. Although the results were not statistically significant in this study, a previous study on the prevalence of *E. bieneusi* in Korean water deer showed higher positivity in the southern region (9/68, 13.2%) than in the central (3/89, 3.4%) and northern (0/24) regions [10]. In this study, differences in the prevalence of *E. bieneusi* by age were not statistically significant. Previous studies also did not find any significant difference according to age, although some have reported conflicting results [16,17]. Therefore, infection is more closely related to individual differences than age [17]. Some of the specimens collected at clinics may have been taken from stray cats that were temporarily sheltered at the clinics or from stray cats that had been adopted by new owners and brought in for treatment. Therefore, further research on the source of infection is necessary.

Four PCR-positive samples were sequenced at Macrogen (Daejeon, Korea), and 2 aligned sequences were obtained using BioEdit 7.2.5. Each sequence was submitted to GenBank (accession numbers PQ534325 and PQ534326). Phylogenetic analysis, including sequence comparisons via NCBI Web BLAST (<http://www.ncbi.nlm.nih.gov/blast>), was also performed. The phylogenetic tree was constructed using MEGA7 [18], and bootstrap analysis was performed using 1,000 replicates. All sequences belonged to group 1, a zoonotic group (Fig. 1). Of the 4 sequences obtained, 3 were Type IV (PQ534326), and one was a new genotype, namely, KCAT1 (PQ534325). The new type, KCAT1, was isolated from a kitten that visited a clinic. The Type IV sequence obtained from humans (AF242478) was used as the reference sequence (243 bp) for genotypic comparison. The KCAT1 genotype exhibited 3 polymorphic sites at nucleotide positions 7 (T to G), 23 (C to T), and 64 (C to T). However, there were no mixed infections of *E. bieneusi* genotypes were observed.

Group 1, the most diverse genotype, has several subgroups (1a–1i) and low host specificity. Along with Group 2, it is zoonotic and has been identified in various animals, including cats [6]. More than 20 *E. bieneusi* genotypes, such as D, Type IV, Peru10, Peru11, and WL11, have been identified in cats and humans [11]. In a previous study on *E. bieneusi* from cats in Korea, only Peru11 (subgroup 1a) was identified [11]. However, the genotypes identified in this study were type IV and KCAT1 genotypes, which are both classified as subgroup 1b/c of Group 1. *E. bieneusi* genotype IV has been identified in humans and in a variety of animals, including cats, dogs, deer, and birds, indicating that this genotype is not restricted to a specific host and has a wide host range. On the contrary, the KCAT1 genotype differs from genotype IV by 3 nucleotides, indicating that it could be a potential transmission route for *E. bieneusi*. However, further studies are necessary to investigate pathogenicity.

Type IV has been identified in cats in several countries, including Colombia, Germany, Portugal, Japan, China, and Türkiye [6,19]. A study in Australia identified a novel *E. bieneusi* genotype in cats, namely, VIC_cat1 (MK696086), which differs from the zoonotic genotype IV by only 1 nucleotide [17]. Type IV, which was previously unidentified in cats in Korea, has been confirmed in cow milk in Korea and is identical to that confirmed in humans (AF242478) [20]. In addition, this Type IV genotype had a 100% sequence match

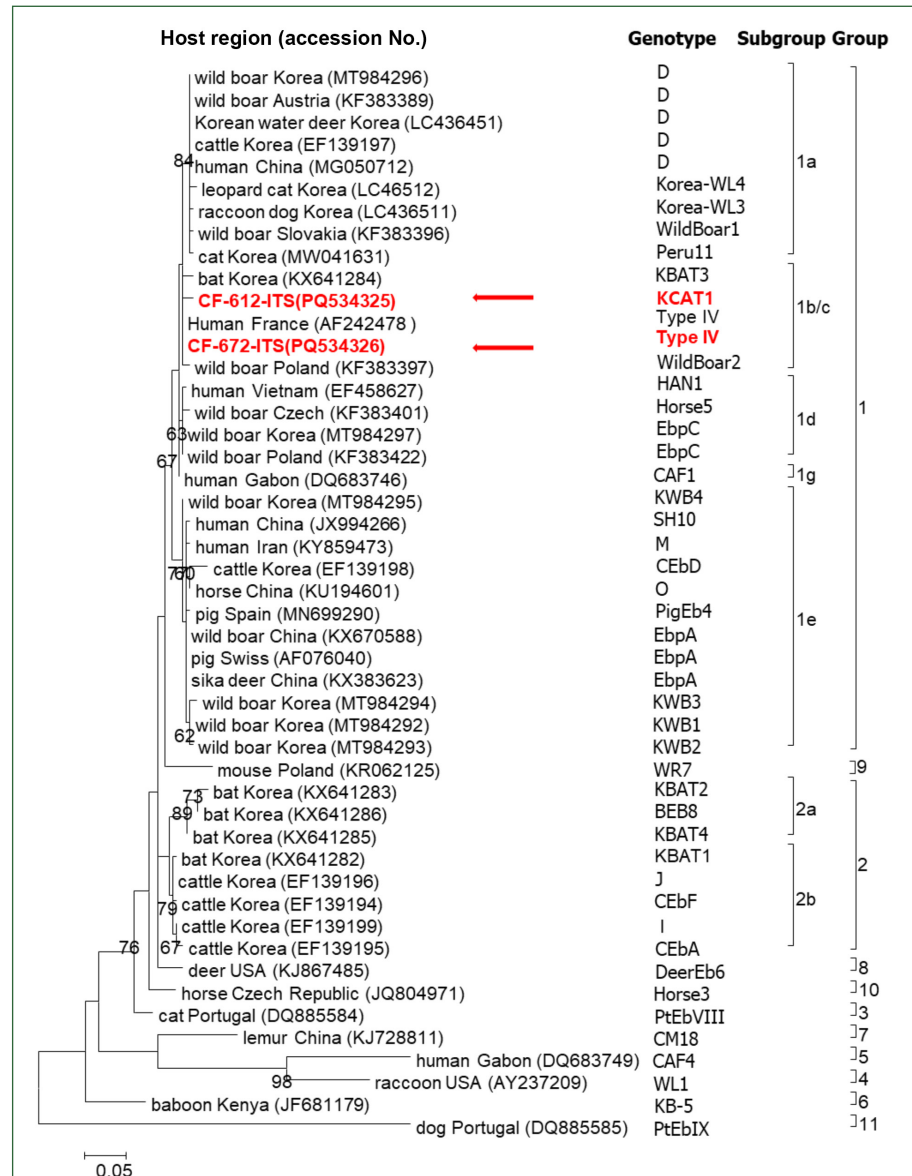


Fig. 1. Analysis of *Enterocytozoon bieneusi* detected in cats in Korea based on the internal transcribed spacer region. The phylogenetic tree was constructed using the maximum likelihood method and bootstrap method with 1,000 replicates in MEGA7. The sequences identified in this study are indicated (arrows). Each sequence is described by host, country, and GenBank accession number. The genotypes (groups and subgroups) are also indicated on the right.

with the human isolate (AF242478).

Dogs and cats are popular companion animals in Korea. They are commonly housed in homes or shelters, which increases their contact with people. In this study, the *E. bieneusi* sequences obtained from cats were 100% identical to those obtained from humans in other countries, indicating that cats represent a potential zoonotic reservoir.

This study is the first nationwide analysis of *E. bieneusi* from cats in Korea, including those from Jeju Island, covering shelter and companion cats visiting clinics. Genotype type

IV, which has not been previously reported in cats in Korea, belongs to Group 1 and exhibits zoonotic potential, along with the new genotype KCAT1. Three of the 4 positive cases were identified in companion cats visiting veterinary clinics; thus, continuous monitoring and further research into the potential zoonotic transmission of *E. bieneusi* are warranted because of their close contact with humans. These findings underscore the importance of routine surveillance of *E. bieneusi* in companion animals. Preventive measures, including enhanced hygiene practices and regular fecal examinations targeting cat owners and veterinarians, are recommended. In addition, the molecular characterization of *E. bieneusi* is essential for a comprehensive understanding of its zoonotic potential, and it plays a critical role in future risk assessments and the development of effective prevention strategies.

References

1. Dashti A, Santín M, Cano L, de Lucio A, Bailo B, et al. Occurrence and genetic diversity of *Enterocytozoon bieneusi* (Microsporidia) in owned and sheltered dogs and cats in Northern Spain. *Parasitol Res* 2019;118(10):2979–2987. <https://doi.org/10.1007/s00436-019-06428-1>
2. Chen M, Wang H, Li X, Guo Y, Lu Y, et al. Molecular epidemiology of *Enterocytozoon bieneusi* from foxes and raccoon dogs in the Henan and Hebei provinces in China. *BMC Vet Res* 2024;20(1):53. <https://doi.org/10.1186/s12917-024-03883-6>
3. Liu X, Wu Y, Yang F, Gong B, Jiang Y, et al. Multilocus sequence typing of *Enterocytozoon bieneusi* isolates from various mammal and bird species and assessment of population structure and substructure. *Front Microbiol* 2020;11:1406. <https://doi.org/10.3389/fmicb.2020.01406>
4. Han B, Pan G, Weiss LM. Microsporidiosis in humans. *Clin Microbiol Rev* 2021;34(4):e0001020. <https://doi.org/10.1128/CMR.00010-20>
5. Nourrisson C, Lavergne RA, Moniot M, Morio F, Poirier P. *Enterocytozoon bieneusi*, a human pathogen. *Emerg Microbes Infect* 2024;13(1):2406276. <https://doi.org/10.1080/22221751.2024.2406276>
6. Li W, Feng Y, Santin M. Host specificity of *Enterocytozoon bieneusi* and public health implications. *Trends Parasitol* 2019;35(6):436–451. <https://doi.org/10.1016/j.pt.2019.04.004>
7. Ruan Y, Xu X, He Q, Li L, Guo J, et al. The largest meta-analysis on the global prevalence of microsporidia in mammals, avian and water provides insights into the epidemic features of these ubiquitous pathogens. *Parasit Vectors* 2021;14(1):186. <https://doi.org/10.1186/s13071-021-04700-x>
8. Lee H, Seo MG, Lee SH, Oem JK, Kim SH, et al. Distribution and genotypic analysis of *Enterocytozoon bieneusi* from wild boars in Korea. *Med Mycol* 2021;59(9):934–938. <https://doi.org/10.1093/mmy/myab030>
9. Lee H, Lee SH, Lee YR, Kim HY, Moon BY, et al. *Enterocytozoon bieneusi* genotypes and infections in the horses in Korea. *Korean J Parasitol* 2021;59(6):639–643. <https://doi.org/10.3347/kjp.2021.59.6.639>
10. Noh G, Lee H, Lee SH, Seo MG, Kim KT, et al. Genotypic analysis of zoonotic *Enterocytozoon bieneusi* in wild deer in Korea. *Parasites Hosts Dis* 2024;62(4):484–489. <https://doi.org/10.3347/PHD.24072>
11. Kwak D, Seo MG. Genetic analysis of zoonotic gastrointestinal protozoa and microsporidia in shelter cats in South Korea. *Pathogens* 2020;9(11):894. <https://doi.org/10.3390/pathogens9110894>
12. Feng S, Jia T, Huang J, Fan Y, Chang H, et al. Identification of *Enterocytozoon bieneusi* and *Cryptosporidium* spp. in farmed wild boars (*Sus scrofa*) in Beijing, China. *Infect Genet Evol* 2020;80:104231. <https://doi.org/10.1016/j.meegid.2020.104231>
13. Zhang XX, Cong W, Lou ZL, Ma JG, Zheng WB, et al. Prevalence, risk factors and multilocus genotyping of *Enterocytozoon bieneusi* in farmed foxes (*Vulpes lagopus*), Northern China. *Parasit Vectors* 2016;9:72. <https://doi.org/10.1186/s13071-016-1356-1>
14. Karim MR, Dong H, Li T, Yu F, Li D, et al. Predominance and new genotypes of *Enterocytozoon bieneusi* in captive nonhuman primates in zoos in China: high genetic diversity and zoonotic significance. *PLoS One* 2015;10(2):e0117991. <https://doi.org/10.1371/journal.pone.0117991>
15. Erkunt Alak S, Can H, Değirmenci Döşkaya A, Sürgeç E, Güvenç M, et al. Molecular prevalence of *Enterocytozoon bieneusi* in stray cats of Izmir, Türkiye. *Comp Immunol Microbiol Infect Dis* 2023;100:102037. <https://doi.org/10.1016/j.cimid.2023.102037>
16. Wang H, Lin X, Sun Y, Qi N, Lv M, et al. Occurrence, risk factors and genotypes of *Enterocytozoon bieneusi* in dogs and cats in Guangzhou, southern China: high genotype diversity and zoonotic concern. *BMC Vet Res* 2020;16(1):201. <https://doi.org/10.1186/s12917-020-02421-4>
17. Zhang Y, Koehler AV, Wang T, Cunliffe D, Gasser RB. *Enterocytozoon bieneusi* genotypes in cats and dogs in Victoria, Australia. *BMC Microbiol* 2019;19(1):183. <https://doi.org/10.1186/s12866-019-1563-y>
18. Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol* 2016;33(7):1870–1874. <https://doi.org/10.1093/molbev/>

msw054

19. Sürgeç E, Güvendi M, Karakavuk M, Erkunt Alak S, Değirmenci Döşkaya A, et al. Genotyping of *Enterocytozoon bieneusi* isolates detected in stray cats of Izmir, Türkiye. *Parasitol Res* 2023;122(11):2729-2735. <https://doi.org/10.1007/s00436-023-07974-5>
20. Lee JH. Molecular detection of *Enterocytozoon bieneusi* and identification of a potentially human-pathogenic genotype in milk. *Appl Environ Microbiol* 2008;74(5):1664-1666. <https://doi.org/10.1128/AEM.02110-07>