



Distribution and genotypes of *Enterocytozoon bieneusi* in raccoon dogs in Korea



Heon-Moo Park¹ , Haeseung Lee² , Su-Jin Chae³ , Kidong Son³ , Sanghyun Lee³ , Kaifa Nazim⁴ , Seung-Hun Lee⁵ , Yoonhoi Koo^{1,6} , Jinsu Kang^{1,6} , Min-Goo Seo^{1,6} , Sang Joon Park^{1,6} , Man Hee Rhee^{1,6} , Dongmi Kwak^{1,6,*}

¹Department of Veterinary Medicine, College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea; ²Veterinary Epidemiology Division, Animal and Plant Quarantine Agency, Gimcheon 39660, Korea; ³Wildlife Disease Research Team, National Institute of Wildlife Disease Control and Prevention, Gwangju 62407, Korea; ⁴Department of Veterinary Parasitology, Khalsa College of Veterinary & Animal Sciences, Punjab 143001, India; ⁵College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea; ⁶Institute for Veterinary Biomedical Science, College of Veterinary Medicine, Kyungpook National University, Daegu 41566, Korea

Abstract

Received: 22 April 2025
Accepted: 22 May 2025

*Correspondence
dmkwak@knu.ac.kr

Citation

Park HM, Lee H, Chae SJ, Son K, Lee S, Nazim K, Lee SH, Koo Y, Kang J, Seo MG, Park SJ, Rhee MH, Kwak D.
Distribution and genotypes of *Enterocytozoon bieneusi* in raccoon dogs in Korea.
Parasites Hosts Dis 2025;63(3):258-263.

Enterocytozoon is a genus of microsporidian parasites, with *Enterocytozoon bieneusi* being a well-known species. It infects various mammalian hosts, including humans, and exhibits zoonotic potential. Out of the 97 fecal and intestinal samples collected from wild raccoon dogs in Korea, 12 (12.4%) tested positive for *E. bieneusi* via PCR, revealing 2 genotypes: genotype D and EbpA. Both genotypes were found to belong to the zoonotic Group 1. Notably, this study is the first to report the EbpA genotype in Korea. Although studies on *E. bieneusi* in raccoon dogs are relatively limited, the findings suggest potential public health concerns.

Keywords: Distribution, genotype, raccoon dogs, *Enterocytozoon bieneusi*, zoonosis

Enterocytozoon is a genus of microsporidian parasites, with *Enterocytozoon bieneusi* being a well-known species that infects a wide range of mammalian hosts, including humans, and exhibits zoonotic potential [1]. The Microsporidia phylum comprises approximately 1,700 identified species, 17 of which are known to infect humans. Among them, *E. bieneusi* accounts for over 90% of the reported human microsporidial infections [2]. Initially identified as an opportunistic pathogen in individuals with acquired immune deficiency syndrome, *E. bieneusi* is now known to cause symptomatic and asymptomatic infections in immunocompromised and immunocompetent individuals [3]. The symptoms typically include diarrhea and lethargy [2].

E. bieneusi is transmitted through the ingestion of spores from contaminated food, and water, or through direct contact with infected sources. Once inside the host, the spores use a polar tube to inject their contents into the epithelial cells of the intestine. The parasite multiplies in the cytoplasm, initially forming multinucleated plasmodia, and then developing into sporogonial stages with complex structures. Sporoblasts form and mature into spores, which are released when the host cell ruptures and are excreted in feces, thereby continuing the transmission cycle [3].

E. bieneusi is primarily detected using molecular and microscopic techniques. PCR, which targets the internal transcribed spacer (ITS) region of the rRNA gene, is considered the most sensitive and specific method. This approach enables detection of isolates and their genotyping for epidemiological studies. To date, over 800 genotypes of *E. bieneusi* have been identified through polymorphism analysis of the ITS region of the rRNA gene

© 2025 The Korean Society for
Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Author contributions

Conceptualization: Kwak D
 Data curation: Park HM, Lee H, Kwak D
 Formal analysis: Park HM, Lee H
 Investigation: Park HM, Lee H, Chae SJ,
 Son K, Lee S
 Methodology: Park HM, Lee H
 Project administration: Kwak D
 Supervision: Kwak D
 Validation: Park HM, Lee H, Chae SJ, Son K,
 Lee S, Nazim K, Lee SH, Koo Y, Kang J,
 Seo MG, Park SJ, Rhee MH
 Visualization: Park HM, Lee H
 Writing – original draft: Park HM, Lee H
 Writing – review and editing: Chae SJ,
 Son K, Lee S, Nazim K, Lee SH, Koo Y, Kang J,
 Seo MG, Park SJ, Rhee MH, Kwak D

Conflict of interest

Dongmi Kwak serves as an editor of *Parasites, Hosts and Diseases* but had no involvement in the decision to publish this article. No other potential conflicts of interest relevant to this study were reported.

ORCID

Heon-Moo Park
<https://orcid.org/0009-0004-4751-4866>
 Haeseung Lee
<https://orcid.org/0000-0002-9587-5488>
 Su-Jin Chae
<https://orcid.org/0000-0002-5147-8843>
 Kidong Son
<https://orcid.org/0009-0001-9930-3743>
 Sanghyun Lee
<https://orcid.org/0009-0005-4687-2909>
 Kaifa Nazim
<https://orcid.org/0000-0002-1524-3035>
 Seung-Hun Lee
<https://orcid.org/0000-0002-6244-0381>
 Yoonhoi Koo
<https://orcid.org/0000-0002-3810-4193>
 Jinsu Kang
<https://orcid.org/0000-0001-5501-7983>
 Min-Goo Seo
<https://orcid.org/0000-0003-1752-5105>
 Sang Joon Park
<https://orcid.org/0000-0003-4275-6060>
 Man Hee Rhee
<https://orcid.org/0000-0002-3088-1318>
 Dongmi Kwak
<https://orcid.org/0000-0003-0876-3179>

[4]. These genotypes have been classified into at least 11 distinct phylogenetic groups based on ITS sequence data [5]. However, recent studies have suggested that these genotypes can be classified into 12–15 groups [5,6]. Group 1 is the largest phylogenetic group and includes numerous genotypes found in humans and animals, such as genotypes D and Type IV, which have been reported in various countries, including Korea [7].

In Korea, *E. bienersi* has been detected in several farm animals, including cattle (genotypes D, Type IV, CEbA, CEbD, CEbF, I, and J), pigs (genotypes PigEBITS3 to PigEBITS5, CAF1, and H), and horses (genotypes horse1 and horse2) [8–10]. It has also been identified in wild animals, such as bats (genotypes KBAT1 to KBAT4, and BEB8), deer, wild boars (genotypes D, H, EbpC, Korea-WL1 to WL2, Korea-WL5 to WL6, KWB1 to KWB4, and RDK), leopard cats (genotype Korea-WL4) [11–13], and shelter cats (genotype Peru11) [14].

Despite these findings, research on the prevalence of *E. bienersi* in raccoon dogs in Korea remains limited. To date, only 1 study has investigated the presence of *E. bienersi* in raccoon dogs and other wild animals in Korea [15]. To address this gap in the literature, this study aimed to investigate the prevalence of *E. bienersi* in wild raccoon dogs in Korea and assess its potential public health significance.

Between May 2017 and October 2023, a total of 97 fecal and intestinal samples were collected from road-killed and trapped wild raccoon dogs across Korea. This study was supported by the Ministry of Environment, Korea. The collection of feces from carcasses was not related to research ethics. Fresh fecal samples were placed in individual tubes, transported in ice-packed containers maintained at 4°C, and delivered to the laboratory for DNA extraction. Relevant sampling information, such as the season and region of collection, was recorded for each sample. Unclear or uncertain data were marked as “unknown.” The sampling regions were categorized based on administrative boundaries into northern (Gangwon and Gyeonggi Provinces) and central/southern (Chungcheong, Gyeongsang, and Jeolla Provinces) areas. DNA extraction was performed using the QIAamp Fast DNA Stool Mini Kit (Qiagen, Hilden, Germany) following the manufacturer’s protocol. The concentration and purity of the extracted DNA were measured using the Infinite 200 PRO NanoQuant plate reader (Tecan, Männedorf, Switzerland). For amplification, PCR was performed using the AccuPower HotStart PCR Premix (Bioneer, Daejeon, Korea), followed by a nested PCR to target and amplify the ITS region of *E. bienersi* for detection purposes [16]. In the first round of PCR, the primers ITSF1 (5′-GGTCATAGGGATGAAGAG-3′) and ITSr1 (5′-TTCGAGTTCTTTTCGCGCTC-3′) were used as the forward and reverse primers, respectively.

In the second round, ITSf2 (5′-GCTCTGAATATCTATGGCT-3′) served as the forward primer and ITSr2 (5′-ATCGCCGACGGATCCAAGTG-3′) as the reverse primer [17]. PCR amplification was performed using the Mastercycler Pro (Eppendorf, Hamburg, Germany). The thermal cycling conditions included an initial denaturation step at 95°C for 5 min, followed by 35 cycles of denaturation at 95°C for 30 sec, annealing at 55°C for 30 sec, and extension at 72°C for 30 sec. A final extension was performed at 75°C for 5 min.

Among the 97 specimens collected, 12 were positive for the *E. bienersi* ITS region (Table 1), resulting in a prevalence of 12.4%. This prevalence was lower than that reported in other animals in Korea, such as wild animals (45.2%), calves (16.9%), cattle (14.9%), and pigs

Table 1. Prevalence of *Enterocytozoon bieneusi* in raccoon dogs in Korea

Variable		Tested	Positive	P-value
Region	Northern	51	4 (7.8)	0.344
	Central and southern	29	5 (17.2)	
	Unknown	17	3 (17.6)	
Season	Spring	24	4 (16.7)	0.070
	Summer	18	2 (11.1)	
	Autumn	25	0	
	Winter	10	3 (30.0)	
	Unknown	20	3 (15.0)	
Total		97	12 (12.4)	

Values are presented as number or number (%).

(14.2%), but higher than that in bats (2.6%) and Korean cats (Jeju Island, 3.8%) [11]. Studies in China reported a prevalence ranging from 2.6% to 28.1%, whereas a study in Poland reported a prevalence of 40.2% [2]. A previous study conducted in Korea considered raccoon dogs as wild animals and reported a prevalence of 35.4% [15]. These studies demonstrate that the prevalence can vary depending on factors such as the geographical region, sample size, and collection period (season or date). Previous studies on raccoon dogs in China have reported variable prevalence rates. In Shandong Province (eastern China), no statistical significance observed based on region, gender, or age, whereas in northern China, significant associations were observed with region and farm scale, but not with the collection year, age, or presence of diarrhea [2,18].

In this study, prevalence of *E. bieneusi* was analyzed by region and season. The northern region had a prevalence of 7.8%, whereas the central and southern regions had a prevalence of 17.2%. By season, the prevalence of *E. bieneusi* was 30.0% in winter, 16.7% in spring, and 11.1% in summer, with no positive cases observed in autumn (Table 1). Statistical analysis using the chi-square test was conducted with the IBM SPSS Statistics version 26 (IBM Corp., Armonk, NY, USA). This analysis showed no statistically significant associations between *E. bieneusi* prevalence and with season or regions, and the association between temperature and region was not clearly established. A clear association between the prevalence of *E. bieneusi* and a specific region could not be confirmed due to the limited number of samples tested for each region and season. This aspect needs to be assessed in follow-up studies.

Four of the 12 PCR-positive samples (2 from feces and 2 from intestinal tissue) were sequenced at MacroGen (Daejeon, Korea). Two aligned sequences were generated using BioEdit version 7.2.5 and submitted to GenBank under the accession numbers PV483842 to PV483845. Phylogenetic analysis, including sequence comparison, was conducted using the NCBI Web BLAST tool (<http://www.ncbi.nlm.nih.gov/blast>).

A phylogenetic tree was constructed using MEGA7, with bootstrap analysis based on 1,000 replicates. All sequences were confirmed to cluster within Group 1, which is known to be zoonotic (Fig. 1). Among the 4 sequences obtained, 3 (RDF-19-ITS, RDS-28-ITS, and RDS-29-ITS) were identified as genotype D (PV483843 to PV483845), whereas 1 (RDF-17-ITS) was classified as genotype Ebpa (PV483842). For genotype comparison and



gesting that raccoon dogs could serve as a potential reservoir for the zoonotic *E. bieneusi* genotype [19,20]. This genotype was also identified in raccoon dogs in Korea [15], including in the current study. The genotype EbpA was first detected in raccoon dogs in a study from China [16]. This genotype has also been identified in humans, with previous studies reporting its presence in various regions and populations [7,20]. However, to date, there has been no evidence of the EbpA genotype in Korea. This study is the first to confirm its presence in the country. These findings underscore the zoonotic potential of the EbpA genotype and its relevance to human health.

Raccoon dogs are among the most commonly found wild animals in Korea observed near urban parks. In this study, the *E. bieneusi* sequences identified in raccoon dogs showed 100% identity with sequences previously reported in humans from other countries, suggesting that raccoon dogs may serve as a potential zoonotic source of infection. This study provides a nationwide analysis of *E. bieneusi* in fecal and intestinal samples from wild raccoon dogs in Korea. Its significance lies in identifying the EbpA genotype, which had not previously been reported in Korea. Like genotype D, EbpA is classified within Group 1 and is considered to have zoonotic potential. Therefore, ongoing research is essential to collect additional samples and expand the geographic scope of surveillance, thereby supporting efforts to monitor the risk of zoonotic transmission through human contact.

Acknowledgments

This study was supported by the National Institute of Wildlife Disease Control and Prevention under a research support project (1800-1833-345).

References

1. Park HM, Lee H, Sung SY, Nazim K, Jang BY, et al. Distribution and genotypic analysis of *Enterocytozoon bieneusi* from cats in Korea. *Parasites Hosts Dis* 2025;63(2):188-194. <https://doi.org/10.3347/PHD.25019>
2. Xue NY, Li ZY, Wang HT, Qin Y, Li XM, et al. High genetic diversity of *Enterocytozoon bieneusi* in minks and raccoon dogs in northern China. *Parasite* 2024;31:71. <https://doi.org/10.1051/parasite/2024071>
3. Nourrisson C, Lavergne RA, Moniot M, Morio F, Poirier P. *Enterocytozoon bieneusi*, a human pathogen. *Emerg Microbes Infect* 2024;13(1):2406276. <https://doi.org/10.1080/22221751.2024.2406276>
4. Chen M, Wang H, Li X, Guo Y, Lu Y, et al. Molecular epidemiology of *Enterocytozoon bieneusi* from foxes and raccoon dogs in the Henan and Hebei provinces in China. *BMC Vet Res* 2024;20(1):53. <https://doi.org/10.1186/s12917-024-03883-6>
5. Jiang S, Yu S, Feng Y, Zhang L, Santin M, et al. Widespread distribution of human-infective *Enterocytozoon bieneusi* genotypes in small rodents in northeast China and phylogeny and zoonotic implications revisited. *Acta Trop* 2024;253:107160. <https://doi.org/10.1016/j.actatropica.2024.107160>
6. Feng K, Yang S, Xu Y, Wen L, Chen J, et al. Molecular characterization of *Cryptosporidium* spp., *Giardia* spp. and *Enterocytozoon bieneusi* in eleven wild rodent species in China: common distribution, extensive genetic diversity and high zoonotic potential. *One Health* 2024;18:100750. <https://doi.org/10.1016/j.onehlt.2024.100750>
7. Li W, Feng Y, Santin M. Host specificity of *Enterocytozoon bieneusi* and public health implications. *Trends Parasitol* 2019;35(6):436-451. <https://doi.org/10.1016/j.pt.2019.04.004>
8. Lee H, Lee SH, Lee YR, Kim HY, Moon BY, et al. *Enterocytozoon bieneusi* genotypes and infections in the horses in Korea. *Korean J Parasitol* 2021;59(6):639-643. <https://doi.org/10.3347/kjp.2021.59.6.639>
9. Jeong DK, Won GY, Park BK, Hur J, You JY, et al. Occurrence and genotypic characteristics of *Enterocytozoon bieneusi* in pigs with diarrhea. *Parasitol Res* 2007;102(1):123-128. <https://doi.org/10.1007/s00436-007-0740-3>
10. Lee JH. Prevalence and molecular characteristics of *Enterocytozoon bieneusi* in cattle in Korea. *Parasitol Res* 2007;101(2):391-396. <https://doi.org/10.1007/s00436-007-0468-0>
11. Lee H, Seo MG, Lee SH, Oem JK, Kim SH, et al. Distribution and

- genotypic analysis of *Enterocytozoon bieneusi* from wild boars in Korea. *Med Mycol* 2021;59(9):934-938. <https://doi.org/10.1093/mmy/myab030>
12. Lee SH, Oem JK, Lee SM, Son K, Jo SD, et al. Molecular detection of *Enterocytozoon bieneusi* from bats in South Korea. *Med Mycol* 2018;56(8):1033-1037. <https://doi.org/10.1093/mmy/myx136>
 13. Noh G, Lee H, Lee SH, Seo MG, Kim KT, et al. Genotypic analysis of zoonotic *Enterocytozoon bieneusi* in wild deer in Korea. *Parasites Hosts Dis* 2024;62(4):484-489. <https://doi.org/10.3347/PHD.24072>
 14. Kwak D, Seo MG. Genetic analysis of zoonotic gastrointestinal protozoa and microsporidia in shelter cats in South Korea. *Pathogens* 2020;9(11):894. <https://doi.org/10.3390/pathogens9110894>
 15. Amer S, Kim S, Han JI, Na KJ. Prevalence and genotypes of *Enterocytozoon bieneusi* in wildlife in Korea: a public health concern. *Parasit Vectors* 2019;12(1):160. <https://doi.org/10.1186/s13071-019-3427-6>
 16. Zhang Y, Xin L, Zhao A, Xu C, Wang T, et al. Molecular detection and genotypes of *Enterocytozoon bieneusi* in farmed mink (*Neovison vison*), blue foxes (*Alopex lagopus*), and raccoon dogs (*Nyctereutes procyonoides*) in Xinjiang, China. *Int J Parasitol Parasites Wildl* 2021;14:211-215. <https://doi.org/10.1016/j.ijppaw.2021.03.003>
 17. Karim MR, Dong H, Li T, Yu F, Li D, et al. Predomination and new genotypes of *Enterocytozoon bieneusi* in captive nonhuman primates in zoos in China: high genetic diversity and zoonotic significance. *PLoS One* 2015;10(2):e0117991. <https://doi.org/10.1371/journal.pone.0117991>
 18. Ma YY, Ma YT, Nie LB, Li TS, Peng JJ, et al. Prevalence and genotype distribution of *Enterocytozoon bieneusi* in farmed raccoon dogs (*Nyctereutes procyonoides*) in Shandong Province, eastern China. *Parasitol Res* 2020;119(6):1873-1878. <https://doi.org/10.1007/s00436-020-06693-5>
 19. Zhao W, Zhang W, Yang Z, Liu A, Zhang L, et al. Genotyping of *Enterocytozoon bieneusi* in farmed blue foxes (*Alopex lagopus*) and raccoon dogs (*Nyctereutes procyonoides*) in China. *PLoS One* 2015;10(11):e0142611. <https://doi.org/10.1371/journal.pone.0142611>
 20. Yang Y, Lin Y, Li Q, Zhang S, Tao W, et al. Widespread presence of human-pathogenic *Enterocytozoon bieneusi* genotype D in farmed foxes (*Vulpes vulpes*) and raccoon dogs (*Nyctereutes procyonoides*) in China: first identification and zoonotic concern. *Parasitol Res* 2015;114(11):4341-4348. <https://doi.org/10.1007/s00436-015-4714-6>